# Concanavalin A (Con A)



9025 Technology Dr. Fishers, IN 46038 • www.bangslabs.com • info@bangslabs.com • 800.387.0672

# INTRODUCTION

Con A is a 104,000 Da lectin (carbohydrate-binding protein) comprised of four identical subunits, and exists as an active dimer or tetramer depending upon pH. Its carbohydrate binding partners are  $\alpha$ -D-glucose and  $\alpha$ -D-mannose with unmodified OH groups at C-3, C-4, and C-6, and terminal glucose residues of proteins and peptides. Con A is a lectin easily isolated from the jack bean, making it a readily available tool for researchers. It has been utilized in a multitude of research projects and diagnostic assays. Cancer research, developing assays for immunoreactions, and studying bacterial mechanisms are only a few examples of how this molecule has been employed by the scientific community.

# **CONJUGATED CON A**

Bangs Laboratories offers Con A magnetic particles as a resource for scientific investigators. Con A microparticles have binding properties similar to the Con A free protein and maintain binding activity. Zem *et. al.* illustrated microspheres linked to Con A can be an alternative to microarrays in the development of carbohydrate-based drugs in diagnostic tests.<sup>17</sup> While Paie *et. al.* demonstrated Con A microspheres prepared by a water-in-oil emulsion technique demonstrated properties of binding to glucose and SAPG-insulin that are similar to the literature values of these properties for unmodified Con A.<sup>11</sup>

Con A-coated BioMag<sup>®</sup>Plus microparticles (BP531) from Bangs Laboratories provide a convenient means for isolating mannosyl- and glucosyl-containing glycoproteins and polysaccharides from serum or cell lysate, or for investigating other lectin / glycan-mediated processes.

### RESOURCES FOR LECTIN RESEARCH

Concanavalin A is one of many types of plant lectins. While we offer Con A conjugated to a magnetic particle, an investigator could also conjugate other types of lectins to polymer, silica, or dyed particles using EDAC covalent immobilization, see *TechNote 205* for more information on conjugation. Table 1 below is a list of resources available for accessing more specific information regarding lectin structure and binding capabilities.

Table 1

Database	Web Link	Specialty
Bacterial carbohydrate structural database	http://www.glyco.ac.ru/bcsdb	The BCSDB scope is "bacterial carbohydrates"
Carbohydrate- active enzymes	http://www.cazy.org/	The database is a dedicated family classification system that correlate with the structure and molecular mechanism of Carbohydrate-active enzymes (CAZymes).
Consortium for Functional Glycomics	http://www.functionalglycomics.org	The Functional Glycomics Gateway is a comprehensive resource for functional glycomics
UniLectin	https://www.unilectin.eu/	An interactive database dedicated to the classification and curation of lectins
Lectins and food	http://poisonousplants.ansci.cornell. edu/toxicagents/lectins.html	A focus on lectin in plants and how it affects livestock
Pathogen—sugar binding database	https://sugarbind.expasy.org/	Website that provides information on known carbohydrate sequences to which pathogenic organisms (bacteria, toxins and viruses) specifically adhere.
Plant lectin database	http://nscdb.bic.physics.iisc.ernet.in/ lectindb/search.html	The lectin database contains structure and sequence information on plant lectins. The database has been completely manually annotated.

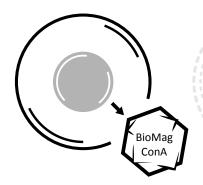
Adapted From: Nilsson, Carol L. "Lectins: Analytical Tools from Nature." Lectins, 2007, pp. 1-13., doi:10.1016/b978-044453077-6/50002-8.

Bangs Laboratories, Inc. Tech Support Doc 0028 Rev. #001, Active: 02/June/2020 << COPY >> Page 1 of 3

# **CONCANAVALIN A AS A RESEARCH TOOL**

The isolation of mannose-containing glycoprotein using Con A affinity chromatography has been a useful tool for investigators.<sup>9</sup> While Protein A chromatography is utilized in many similar types of separations, it is not a good candidate for large-scale purification because of its high cost<sup>3</sup>; consequently, many assays have been built around Con A.

Due to its ability to bind mannosyl- and glucosyl-containing glycoprotein present in mammalian cells, agglutination assays have been developed. These assays have been adapted for investigating tumors in canines<sup>4</sup>, rats<sup>7</sup>, and human systems<sup>15</sup>. More recently, assays have been developed using Con A as an agent to test immune responsiveness<sup>5</sup>. BioMag<sup>®</sup> Concanavalin A is used to adhere magnetic particles to cell nuclei for CUT&RUN, a chromatin profiling protocol that has several key advantages over chromatin immunoprecipitation (ChIP). ChIP has low efficiency due to the millions of cells required, high background from tens of millions of reads as a result of sonication, and low resolution by reading hundreds of base pairs as opposed to possible single base pair resolution, see references for more details.









BioMag ConA immobilzes cell nucleus

Transcription Factor Ab diffuses out

ProA-MNase diffuses in

Add Ca<sup>2</sup> { start reaction } - Chelator { stop reaction } (liberating target sequence)

Con A application is not used exclusively in mammalian systems. The properties of Con A have been utilized to probe the characteristics of microbes as well. By blocking glycoprotein receptors on cell surfaces, bacterial phenotypes can be explored with regards to receptor binding. It has been utilized in the study of bacterial cell-wall structure<sup>6</sup>, investigations of bacterial attachment mechanisms<sup>10</sup>, slime molds surface receptors<sup>16</sup>, and virus interactions with mammalian cells<sup>8</sup>.

# **REFERENCES**

- 1. Ambrosi, M., Cameron, N. R., & Davis, B. G. (2005). *Lectins: Tools for the Molecular Understanding of the Glycocode*. ChemInform, 36(35). doi: 10.1002/chin.200535314
- 2. Becker, F. F. (1975). Concanavalin A Agglutination of Cells from Primary Hepatocellular Carcinomas and Hepatic Nodules Induced by N-2-Fluorenylacetamide. Cancer Research, 35, 2879–2883.
- 3. Bereli, N., Akgöl, S., Yavuz, H., & Denizli, A. (2005). *Antibody purification by concanavalin A affinity chromatography*. Journal of Applied Polymer Science, 97(3), 1202–1208. doi: 10.1002/app.21862
- 4. Betton, G. R. (1976). Agglutination reactions of spontaneous canine tumor cells, induced by concanavalin A, demonstrated by an isotopic assay. International Journal of Cancer, 18(5), 687–696. doi: 10.1002/ijc.2910180518
- 5. Bílková, B., Albrecht, T., Chudíčková, M., Holáň, V., Piálek, J., & Vinkler, M. (2016). *Application of Concanavalin A during immune responsiveness skin-swelling tests facilitates measurement interpretation in mammalian ecology.* Ecology and Evolution, 6(13), 4551–4564. doi: 10.1002/ece3.2211
- 6. Doyle, R. J., & Birdsell, D. C. (1972). *Interaction of Concanavalin A with the Cell Wall of Bacillus subtilis*. Journal of Bacteriology, 109(2), 652–658. doi: 10.1128/jb.109.2.652-658.1972
- 7. Kakizoe, T. (1980). *Increased Agglutinability of Bladder Cells by Concanavalin A after Administration of Carcinogens*. Cancer Reasearch, 40, 2006–2009.
- 8. Leon, M. P. D., Hessle, H., & Cohen, G. H. (1973). *Separation of Herpes Simplex Virus-Induced Antigens by Concanavalin A Affinity Chromatography.*Journal of Virology, 12(4), 766–774. doi: 10.1128/jvi.12.4.766-774.1973
- 9. Litman, B. J. (1982). [23] *Purification of rhodopsin by concanavalin A affinity chromatography. Methods in Enzymology Biomembranes* Part H: Visual Pigments and Purple Membranes I, 150–153. doi: 10.1016/s0076-6879(82)81025-2
- 10. Ofek, I., Mirelman, D., & Sharon, N. (1977). Adherence of Escherichia coli to human mucosal cells mediated by mannose receptors. Nature, 265(5595), 623–625. doi: 10.1038/265623a0

Bangs Laboratories, Inc. Tech Support Doc 0028 Rev. #001, Active: 02/June/2020 << COPY >> Page 2 of 3

- 11. Pai, C. M., Bae, Y. H., Mack, E. J., Wilson, D. E., & Kim, S. W. (1992). *Concanavalin a Microspheres for a Self-Regulating Insulin Delivery System.*Journal of Pharmaceutical Sciences, 81(6), 532–536. doi: 10.1002/jps.2600810612
- 12. Saleemuddin, M., & Husain, Q. (1991). *Concanavalin A: A useful ligand for glycoenzyme immobilization—A review.* Enzyme and Microbial Technology, 13(4), 290–295. doi: 10.1016/0141-0229(91)90146-2
- 13. Sela, B.-A., Lis, H., Sharon, N., & Sachs, L. (1971). *Quantitation of N-acetyl-d-galactosamine-like sites on the surface membrane of normal and transformed mammalian cells*. Biochimica Et Biophysica Acta (BBA) Biomembranes, 249(2), 564–568. doi: 10.1016/0005-2736(71)90132-5
- 14. Sparbier, K. (2005). Selective Isolation of Glycoproteins and Glycopeptides for MALDI- TOF MS Detection Supported by Magnetic Particles. Journal of Biomolecular Techniques, 16(4), 407–413.
- 15. Voyles, B. A., & Mcgrath, C. M. (1976). *Markers to distinguish normal and neoplastic mammary epithelial cells in vitro: comparison of saturation density, morphology and concanavalin a reactivity.* International Journal of Cancer, 18(4), 498–509. doi: 10.1002/ijc.2910180415
- 16. West, C. M., & Mcmahon, D. (1977). *Identification of concanavalin A receptors and galactose-binding proteins in purified plasma membranes of Dictyostelium discoideum.* The Journal of Cell Biology, 74(1), 264–273. doi: 10.1083/jcb.74.1.264
- 17. Zem, Gregory C., et. al. "Microbead Analysis of Cell Binding to Immobilized Lectin: an Alternative to Microarrays in the Development of Carbohydrate Drugs and Diagnostic Tests." Acta Histochemica, vol. 108, no. 4, 2006, pp. 311–317., doi:10.1016/j.acthis.2006.03.019.

Bangs Laboratories, Inc. Tech Support Doc 0028 Rev. #001, Active: 02/June/2020 << COPY >> Page 3 of 3