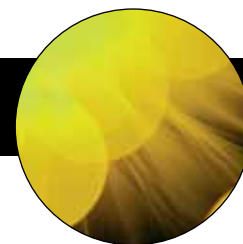


# Microsphere Selection

## Which Beads are Best for Your Application?



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### Suggestions and strategies for choosing the best beads for common microsphere applications.

Microspheres offer a highly convenient and flexible system for developing reagents for assays and bioseparations, and for use as instrument standards. With many varieties of microspheres available, it is important to think about the demands the application will place on them when choosing a base bead. Physical and optical properties should be considered in the context of handling and detection, and thought should also be given to requirements for diameter and size distribution, composition, surface chemistry, and any other needed properties.

#### Diameter

Microsphere size may be critical to the proper function of an assay, or it may be a secondary consideration. Considering traditional diagnostic methods, the test or assay format commonly dictates particle size, such as the use of very small spheres ( $\sim 0.1$ - $0.4\mu\text{m}$ ) in lateral flow tests, or the use of cell-sized spheres ( $\sim 4$ - $10\mu\text{m}$ ) for bead-based flow cytometric assays. In magnetic separations, the exact size of the magnetic particle may be unimportant provided that the particles are in some general size range, and offer desired separation characteristics.

Diameter also determines surface area. Small-diameter spheres present more surface area per unit weight, while larger spheres present more surface area per bead. Size also affects ease of handling, processing considerations, and the amount of reagent needed for coating.

#### Composition

Common microsphere compositions include polystyrene (PS), poly(methyl methacrylate) (PMMA), and silica. These materials possess different physical and optical properties, which may present advantages or limitations for different applications.

Polymer beads are generally hydrophobic, and as such, have high protein binding abilities. However, they often require the use of some surfactant in the storage buffer to ensure ease of handling.

During synthesis, functional monomers may be copolymerized with styrene or methyl methacrylate to develop beads with surface reactive groups. Functional groups may be used in covalent binding reactions, and aid in stabilizing the suspension.

Silica microspheres are inherently hydrophilic and negatively charged. Consequently, aqueous silica suspensions rarely require use of surfactants or other stabilizers. Carboxyl- and amine-functionalized silica microspheres are available for use in common covalent coating protocols, and plain silica spheres may be modified using a variety of silanes to generate functional groups or alter surface properties.

#### Coating

Microspheres may be coated with capture molecules, such as antibodies or oligonucleotides, for use in diagnostic or separation applications. Microsphere coatings are typically optimized to achieve desired specific activity, while minimizing nonspecific interactions. Consideration should be given to the required stability, development time frame and budget, and the specific biomolecule to be coated. These factors will help determine the most fitting coating strategy (adsorption, covalent coupling, or affinity binding) for short- and long-term objectives.

#### Adsorption

Adsorption relies primarily on hydrophobic interactions between the biomolecule and polymer particle. Such coatings are fairly simple to conduct, involving incubation of the microspheres with the purified biomolecule. They typically require little optimization, and reagents may be developed relatively quickly. However, as adsorption relies on the formation of multiple attachment points between the molecule and particle, this strategy is typically reserved for use with proteins and non-functionalized polymer spheres. Adsorption is generally not suitable for hormones, peptides, or nucleic acids in hybridization-based applications, and protein adsorption to silica is expected to be less efficient than to polymer.

## Covalent Coupling

Covalent coupling results in the permanent attachment of the molecule to the functionalized microsphere. It can provide needed stability when developing a commercial reagent, and for multiplexed assays, where analyte-specific bead populations are mixed. Additionally, specialized chemical linkers may be employed to address steric effects or to optimally orient the molecule. Although covalent binding protocols often involve a higher level of optimization than other approaches, coupling kits are available to simplify the process.

## Affinity Binding

Affinity binding is a straightforward method for immobilizing primary antibodies or biotinylated molecules. Proteins A and G and Fc-specific antibody coatings permit the directed immobilization of primary antibodies, and streptavidin is used extensively for the binding of biotinylated molecules, such as antibodies, peptides, and oligonucleotides.

Test / Assay Format	Bead Size	Bead Type	Coating Strategy
Flow cytometric (suspension array)	2 – 15µm	QuantumPlex™ QuantumPlex™™ (for multiplexing) or Non-fluorescent (simplex or multiplex with different bead sizes)	Covalent or streptavidin / biotin
Lateral Flow	0.1 – 0.4µm	Dyed (visible or fluorescent)	Covalent or adsorption
Lateral Flow – Boulders in the Stream	0.1 – 0.4µm mobile phase	Dyed (visible) mobile phase	Covalent or adsorption
	~2 – 3µm capture phase	Undyed capture beads	
Dipstick	0.1 – 0.4µm	Dyed (visible)	Covalent or adsorption
LAT ("Latex" Agglutination Test)	0.2 – 1.0µm	Undyed or visibly dyed	Covalent or adsorption
Light Scattering (Automated LAT– Nephelometric or Turbidimetric)	0.36– 0.76µm (diameter equal to wavelength of light being scattered)	Undyed	Covalent

It is important to note that each binding strategy has benefits and limitations, which should be weighed in the context of study objectives and the demands that will be placed on the finished reagent.

## Special Properties

Many applications in the life sciences demand added properties, such as fluorescence or a visible color, or iron oxide inclusions for magnetic separations. Polymer spheres (and polymer-based magnetic spheres) are often internally dyed via organic solvent swelling, and many standard products are available. Dye concentrations can be adjusted to produce beads with different intensities to meet special needs, such as QuantumPlex™ for multiplexed flow

cytometric assays, or our Dragon Green or Flash Red Intensity Standards, which support imaging applications and associated instrument QC. Many surface- or internally-labeled fluorescent beads are also available as specialized flow cytometry standards.

Various types of superparamagnetic microparticles are available as well – with different matrices, magnetite content, surface groups, etc. For new assays or applications, magnetic beads should be evaluated with application demands in mind.

Separation	
Cells	BioMag® anti-CD marker or secondary antibody
Subcellular organelles	BioMag®
Immunoprecipitates	ProMag™ Protein G, BioMag® secondary antibody, or Protein A or G
mRNA	BioMag® Oligo dT (20) or mRNA Purification System
Biotinylated oligonucleotide capture or binding	ProMag™, COMPEL™, or BioMag® Streptavidin
Biopanning	ProMag™, COMPEL™, or BioMag®
Assay	
Immunoassays	ProMag™, COMPEL™, or BioMag®
Hybridization-based assays	ProMag™ or COMPEL™
Flow cytometric assays	COMPEL™

For additional information regarding microsphere selection, visit our website at [www.bangslabs.com](http://www.bangslabs.com).



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We also stand behind our products. Regardless of the size of your question or the size of your company, we offer tech support, absolutely free.

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