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**B E A D S ● A B O V E T H E R E S T™**

## Description

Though many of our microsphere suspensions contain an antimicrobial agent, certain applications require sterile preparations. In these instances, suspensions may be treated with radiation, heat, or chemicals to achieve a microbe-free status.

## Procedure

Researchers are advised to optimize the use of particles in any application, and to validate the decontamination procedure.

### Gamma Irradiation

A dose of 0.030 megarad/hour for 24 hours (0.72 megarad) is sufficient to control most bacteria and contaminating molds; however, a few yeast cells may survive beyond 100 hours (3.0 megarads). Irradiation of particles after packaging at >2 megarads (67 hours) gives excellent results. In some cases, the radiation will cause discoloration of the packaging or certain soluble materials, but the particles seem to be unaffected. This process, if available, is quite effective for sterilizing the particles.

### Heat Treatment of Aqueous Suspensions

This applies to aqueous suspensions in high density polyethylene bottles and closures.

1. Preheat an appropriately sized, calibrated oven to  $80^{\circ}\text{C} \pm 4^{\circ}\text{C}$ .
2. Place bottles into preheated  $80^{\circ}\text{C}$  oven. Each bottle may touch another bottle, but may not touch the walls or top of the oven. Only the bottom of the bottles may come in contact with the oven surface.
3. Allow the material to remain in the oven for 2 hours  $\pm$  10 minutes.
4. Remove from  $80^{\circ}\text{C}$  oven and incubate in a preheated  $40^{\circ}\text{C} \pm 4^{\circ}\text{C}$  calibrated oven for 20 hours  $\pm$  4 hours. The same oven may be used for this temperature cycling if the temperature is reduced from  $80^{\circ}\text{C}$  to  $40^{\circ}\text{C}$  in one hour.
5. Remove from  $40^{\circ}\text{C}$  oven and incubate in  $80^{\circ}\text{C} \pm 4^{\circ}\text{C}$  calibrated oven for 2 hours  $\pm$  10 minutes. The same oven may be used for this temperature cycling if the temperature is increased from  $40^{\circ}\text{C}$  to  $80^{\circ}\text{C}$  in one hour.
6. Remove from  $80^{\circ}\text{C}$  oven and incubate in a preheated  $40^{\circ}\text{C} \pm 4^{\circ}\text{C}$  calibrated oven for 20 hours  $\pm$  4 hours. The same oven may be used for this temperature cycling if the temperature is reduced from  $80^{\circ}\text{C}$  to  $40^{\circ}\text{C}$  in one hour.
7. Remove from  $40^{\circ}\text{C}$  oven and incubate in  $80^{\circ}\text{C} \pm 4^{\circ}\text{C}$  calibrated oven for 2 hours  $\pm$  10 minutes. The same oven may be used for this temperature cycling if the temperature is increased from  $40^{\circ}\text{C}$  to  $80^{\circ}\text{C}$  in one hour.
8. In total, the product should be exposed to three cycles of  $80^{\circ}\text{C}$  and two cycles of  $40^{\circ}\text{C}$ .

### Sterilization by Rinsing with 70% Ethanol or 70% Isopropyl Alcohol

#### Notes:

- This procedure should only be used with undyed, uncolored polymer or silica microspheres. Exposure of colored beads to ethanol or isopropyl alcohol may leach out the dye.
- Particles that are  $0.5\mu\text{m}$  or larger in diameter should be used as they will be able to form a pellet by centrifugation. For particles smaller than  $0.5\mu\text{m}$ , a hollow fiber filter or dialysis tubing must be used to concentrate the particles.
- Size standard particles should not be exposed to ethanol or isopropyl alcohol because it may temporarily swell the particles. They will return to their approximate size once resuspended in water, but their accuracy as size standards may be compromised. Use an alternate method for sterilization.

1. Mix the bottle of particles by inverting the bottle several times to achieve an even distribution of the particles before taking an aliquot.
2. Place an aliquot of the bead suspension into a centrifuge tube. Centrifuge the particles to form a visible, white pellet at the bottom

of the tube following the instructions on the next page.

3. Remove the water supernatant and replace with an equivalent volume of 70% ethanol or 70% isopropyl alcohol.
4. Vortex briefly to mix, then centrifuge down to pellet and remove the ethanol or isopropyl alcohol, replacing with fresh 70% ethanol or 70% isopropyl alcohol. Repeat this two times.
5. After the last 70% ethanol or 70% isopropyl alcohol rinse, centrifuge to form a pellet and resuspend in an equivalent amount of sterile DI water or the desired sterile aqueous buffer. Vortex briefly to mix.
6. Centrifuge to form a pellet, remove supernatant, and replace with fresh sterile DI water or sterile aqueous buffer.
7. Repeat the sterile DI water or sterile buffer wash two more times to remove all traces of ethanol or isopropyl alcohol before using the particles. Particles should now be ready to use.

### Centrifugation

Washing particles may be done via centrifugation. This procedure must be performed carefully. Excess centrifugation will result in resuspension difficulties. For the purposes of pelleting, it is important to understand the settling velocities of particles.

For spherical particles, settling velocity can be calculated using Stokes' Law. (See also TechNote 206 for a reference table and additional equations.)

$$V = \frac{2ga^2(\rho_1 - \rho_2)}{9n}$$

- V = Velocity in cm/sec
- g = g force in cm/sec<sup>2</sup>
- $\rho_1$  = density of particle in g/cm<sup>3</sup>
- $\rho_2$  = density of suspending media in g/cm<sup>3</sup>
- n = coefficient of viscosity in poises (g/cm-sec)
- a = radius of spherical particle in cm

For calculating the settling velocity of polystyrene microspheres at 1G in 20°C water, Stokes' Law can be expressed in the following formula, where d = diameter in microns,  $\rho_1 = 1.05 \text{ g/cm}^3$ ,  $\rho_2 = 1.00 \text{ g/cm}^3$ , and  $n = 1.002 \text{ cp}$ .

$$V = 2.77 \times 10^{-6}d^2$$

To estimate appropriate times for centrifugation, settling velocity is multiplied by the G forces generated by the centrifuge. The resultant velocity is then compared to the height of the centrifuge tube.

*For example:* A 1.0µm particle placed in a microcentrifuge generating 10,000 G will settle at a velocity of  $2.77 \times 10^{-2} \text{ cm/sec}$ . Pelleting the particle in a 4cm high tube would require a 144 second (minimum) centrifuge run. The actual time required to form an acceptable pellet could possibly be 50% longer. These calculations are intended to be used as guidelines to assist in determining centrifugation time. Different size particles yield dramatically different settling velocities. A 10.0µm particle could settle in 2 seconds under the aforementioned conditions, whereas a 0.01µm particle could take at least 4 hours to settle. Brownian motion and particle concentration also affect settling rates.

### Storage and Stability

Store at 2-8°C. Freezing of particles may result in irreversible aggregation and loss of binding activity.

**This product is for research use only and is not intended for use in humans or for *in vitro* diagnostic use.**

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